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Synthesis and cytotoxicity of (+/–)-7,9-dideoxypancratistatin analogues†

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Access to some 7,9-dideoxy analogues of pancratistatin was achieved by combining our previously reported nitroenal/dioxanone annulation (to form ring C) with a hetero-Diels–Alder/aromatization path to build the dihydroisoquinolinone subunit (rings A and B); testing of their antiproliferative activity afforded some clues about the role of aromatic substituents in pancratistatin's pharmacophore.

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Introduction

The isocarbostyril constituents of the Amaryllidaceae plant family, with general structure **A** (Fig. 1),¹ keep receiving considerable attention due to their antitumoral properties.² Pancratistatin (1),³ in particular, is at the focus of much research activity⁴ because it selectively induces apoptosis in a wide variety of human cancer cell lines at low doses with minimal effect on normal cells,⁵ it decreases the volume of human tumors *in vivo*⁶ and does not inhibit CYP3A4.⁷ A major and continuous effort is being dedicated to the total synthesis of **1** with the main goal of solving the supply problem derived from its minute occurrence in natural sources.^{8,9} The synthetic work is also being directed to the preparation of unnatural derivatives to improve the pharmacological profile (in particular, to gain higher water solubility) and to define the pharmacophore.

To date, a significant number of pancratistatin derivatives have been prepared and tested.¹⁰ Most of them have changes in rings B and C. In general, any departure from the B/C natural structure usually resulted in diminished activity. In fact, a number of structural elements were identified as critical motifs to be preserved for potent activity. They include the location and the stereochemistry of three hydroxyl groups in ring C (those at carbons C2, C3 and C4),¹¹ the *trans* B/C ring junction (positions C4 and C10b)¹² and the ring B-lactam functionality (positions C5 and C6).¹³ In clear contrast, position C1 was demonstrated to be the only one that could be



Fig. 1 A: general structure for the isocarbostyril constituents of the Amaryllidaceae family; ^a the C3-absolute configuration is as shown except for natural 3-*epi*-pancratistatin. Compounds **1**, **2**, **4** and **5**: hitherto known ring-A substitution patterns for the B/C amide–cyclohexanetetrol pancratistatin system [in natural (**1**, **2**) and synthetic (**4**, **5**) derivatives]. Compound **3**: indole mimic of **2**.

altered with advantage. This line of research, first initiated with the C1-benzoate of pancratistatin that showed $\approx 10^3$ increments in antitumoral activities,¹⁴ continues to be successfully pursued by several groups.¹⁵ The participation of ring A in the pharmacophore has been much less studied. The phenolic C7-group is important for activity, as early established with the comparatively less active natural 7-deoxy analogues [*e.g.*, 7-deoxypancratistatin in lung NCI-H460 and colon KM20L2 human cancer cell lines].¹⁷ Thus far, only three other derivatives of pancratistatin with variations in ring A (while keeping intact its B/C functionality) were synthesized: the indole mimic of 7-deoxypancratistatin **3**,¹⁸ the bisTMS derivative **4**¹⁹ and the

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Part I. The Annulation + Electrophilic Aromatic Substitution (EAS) pathway to pancratistatins



7,8-deoxy-9-methoxypancratistatin 5^{20} (Scheme 1). Replacement of the highly oxygenated phenyl ring A with a plain indole unit as in 3 resulted in the drop of activity against a number of human cancer lines to GI₅₀ values higher than 10 µg mL⁻¹. Complete deoxygenation of ring A as in 4 (with the incorporation of two trimethylsilyl groups in positions 8 and 9) also caused loss of activity. Interestingly, the presence of a single oxygenated substituent at position C9 in ring A, as it happens in the methoxy analogue 5, can be enough to keep the activity in the micromolar range (GI₅₀ = 2.6–4.9 µg mL⁻¹).

Further understanding of the role that ring A plays in the pharmacophore requires the synthesis of new derivatives with different aromatic substituents. To that end, the development of preparative methods that could eventually allow the access to diverse substitution patterns through common intermediates is most desirable.

We recently reported a synthetic protocol to the pancratistatin skeleton (Scheme 1, part I)²¹ in which ring C is assembled through a formal [3 + 3] annulation of β -aryl- α -nitro- α , β -enals (8)²² with 2,2-dimethyl-1,3-dioxan-5-one (8 + 9 \rightarrow 10), and ring B is formed by intramolecular electrophilic aromatic substitution (EAS) of appropriately-protected methyl carbamates of type 11. In this approach, the substituents present in the starting aldehydes **6** and the activation-requirements and regioselectivity of the EAS step dictate and restrict the attainable substitution pattern of ring A in the final products **12**.

Looking for more flexible pathways, we identified compounds of type **16** as attractive intermediates because their unsaturated furylamine or furylamide functionality could in principle be transformed into differently-substituted dihydroisoquinolin-1(2*H*)-ones (the A/B ring system of **12**) by a number of procedures. In particular, we set out to explore the two paths, a and b, indicated in part **II** of Scheme 1. In path a, a furan–alkyne gold-catalysed cycloisomerization²³ of **16** would directly afford phenolic tetrahydroisoquinolines **17** (R⁷ or R' = OH), which should then be oxidized at their benzylic position and N-deprotected to render the corresponding dihydroisoquinolinone units. In path b, **16** would have to undergo an intramolecular Diels–Alder cycloaddition of its furan ring with the *N*-tethered unsaturated chain and the resulting oxabicyclic systems **18** would then need to be aromatized.

Herein, we report an initial study of both paths, the first successful implementation of path b and its application to the preparation of (+/-)-7,9-dideoxy derivatives of pancratistatin, which were tested against the NCI-H460 and the MCF-7 tumoral cell lines.



Scheme 2 Synthesis of furylpropargylamines 16a and 16b and attempts of gold-catalysed cycloisomerization to phenols 17.

Exploratory studies of the gold-catalysed cycloisomerization pathway

All intermediates of type **16** used in this work were synthesized from the furyl-derived nitrocyclitol (+/–)-**15**, in turn prepared as previously reported²⁴ in three steps from furfural and dioxanone **9** through the β -furyl- α -nitroenal **14** (Scheme 1, part II). In particular, as indicated in Scheme 2, the application of two different NO₂-reduction and protection protocols to **15** led to intermediates **19** and **20**, which were respectively converted into the desired furylpropargylamines **16a** and **16b** by incorporation of a propargyl group at their nitrogen atoms.

Unfortunately, all our attempts to promote the gold-catalysed cycloisomerization of either the cyclic-carbamate protected propargylamine **16a** or its less-rigid *N*-tosyl analogue **16b** under a variety of conditions, using AuCl₃, $[\mu$ -Cl(AuPPh₃)₂]-BF₄ or salt **21**, failed thus far to render the desired phenolic tetrahydroisoquinolines **17**; either non-reaction/deprotection or complex mixtures were observed.

The hetero-Diels-Alder/aromatization pathway

To explore path b, we selected acrylamide **16d** (Scheme 3). We chose to use a chlorine substituent next to the oxygen



Scheme 3 Incorporation of the IMDAF/aromatization protocol to dihydroisoquinolin-1(2*H*)-ones into a new synthetic scheme for 7,9-dideoxy pancratistatin analogues (+/–)-**12** from furfural.

atom of its furan ring with a double purpose; first, to facilitate the IMDAF cycloaddition (*i.e.*, the intramolecular Diels–Alder reaction of furan, step 1b in path b, Scheme 1);²⁵ second, to enable the aromatization process (step 2b) to directly afford C8-phenolic-7,9-dideoxygenated isoquinolinone systems, which we judged useful for pharmacophore studies (*vide infra*).

For the synthesis of **16d** from (+/-)-**15**, we first addressed the reduction of the nitro group, which was best performed with RANEY®-Nickel on its methoxyisopropyl ether derivative **22** (Scheme 3). We then introduced an acryloyl group at the nitrogen atom of the resulting amine **23**, replaced the acetal function at C4 with a methylcarbamate, and finally chlorinated the furan ring.

Acrylamide **16d** showed no change on heating in toluene in a closed flask at 120 °C for 3 days, but decomposed at higher temperature (xylenes, 130–145 °C), or when smoothly warmed (60–90 °C) in the presence of Yb(OTf)₃. However, when it was heated in the presence of NaHCO₃ (closed flask, toluene or xylenes, external bath temperature = 130–145 °C), the desired cycloadduct **18a** was obtained as a single stereoisomer (30–53%, Scheme 3).²⁶

The conversion of **16d** into **18a** is the first example of an IMDAF process of a *secondary* amide *N*-tethered to a 2-furyl ring to directly render the corresponding *N*-deprotected oxabicyclic-fused γ -lactam.²⁷

Once the viability of the IMDAF process was demonstrated, we were ready to study the following steps in the way to **12**, *i.e.*, the conversion of cycloadduct **18a** into the corresponding pancratistatin analogues. In practice, we found it convenient to first elaborate ring C before proceeding to the opening-aromatization of the oxanorbornene unit. In the event, successive treatment of **18a** with acid (to cleave the acetonide), Na(AcO)₃BH (to reduce the keto group from the β face) and Ac₂O (steps 8–10) led to triacetate **18b** (85% overall) with the natural relative configuration at every stereocenter of ring C.

Aromatization of **18b** was simply achieved with NaMeO, which also hydrolysed the ester and carbamate groups, rendering the desired 7,9-dideoxy-8-phenolic pancratistatin analogue **12a**. By inverting the order of the reducing and deprotection steps in the synthetic sequence (steps a–d at the bottom of Scheme 3), **18a** was converted into **12c**, the C2-epimer of **12a**. We also prepared the analogue **12b** by alkylation of **12a** with benzyl bromoacetate.

Antiproliferative activity

The antiproliferative data of compounds (+/–)-**12a–c** against the tumoral cell line NCI-H460 (human large-cell lung carcinoma) are collected in Table 1. The activity data for some reference compounds are also included.

As compared to (+/–)-7-deoxypancratistatin [(+/–)-2], which showed an NCI-H460 cell growth inhibition (%GI) of 85% (at 100 μ M) and an IC₅₀ value of 1.57 μ g mL⁻¹,²⁸ its 9-deoxy-8-

hydroxy analogue **12a** as well as its 9-deoxy-8-[2-(benzyloxy)-2oxoethoxy] analogue **12b** displayed %GI values of only 3.8% and 7% (respectively, at the same concentration) and IC₅₀ values higher than 30 µg mL⁻¹. Thus, removing the oxygenated substituent at C9 is extremely detrimental to activity (>12 fold), regardless of whether the remaining C8 oxygenated function is a free phenol, as in **12a**, or an alkylated form of it, as in **12b**. This decrease in activity is by far more pronounced than those reported for the removal of the oxygenated function at C7 [a \approx 6 fold drop in going from pancratistatin (**1**, IC₅₀ = 0.048 µg mL⁻¹) to its 7-deoxy analogue **2** (IC₅₀ = 0.29 µg mL⁻¹)], or at C8 [a \approx 10 fold cut in going from 7-deoxypancratistatin (**2**, IC₅₀ = 0.29 µg mL⁻¹) to its 8-deoxy-9-methoxy analogue **5** (IC₅₀ = 2.8 µg mL⁻¹)].

Thus, of the three oxygenated substituents that natural pancratistatin has in its aromatic ring A, the one at C9 appears to be the most important for activity. Its location, at position *para* with respect to the amide carbonyl function, suggests that it could act, independently of other functions, by tuning the acceptor capabilities of the mentioned carbonyl group through a donor resonance effect.

Removal of the oxygenated function in C9 was also adverse in the (comparatively inactive) 2-*epi* series: the activity dropped in going from (+/–)-7-deoxy-2-*epi*-pancratistatin [(+/–)-2-*epi*-2, %GI = 16%] to its 9-deoxy-8-hydroxy analogue **12c** (%GI = 3.5%).

Similar data were obtained against the MCF-7 (breast) tumoral cell line: **12a** and **12b** showed %GI values of 4.6% and 16%, respectively, while **12c** was found to be completely inactive (for details, see ESI[†]).

Conclusions

The capability of compounds of type **16** to serve as intermediates for the preparation of pancratistatin analogues was initially evaluated by exploring two different paths to build the A/B ring system. In spite of initial failure, the furan–alkyne gold-catalysed cycloisomerization route (path a) deserves to be further studied because of its high potential. The Diels–Alder/ aromatization pathway (path b) was reduced to practice and successfully incorporated into a new synthetic scheme for pancratistatin analogues from furfural. Because the IMDAF process takes place with *secondary* acrylamides of type **16** (NPG = NH), the synthetic protocol does not require protection–deprotection steps at nitrogen. Testing of the cytotoxic activities of the 7,9-dideoxy analogues allowed further

Table 1 Antiproliferative data for 12a-c and some reference compounds against the tumoral cell line NCI-H460 (large-cell lung human carcinoma)

	(+/-)-2 ^a	(+/-)- 12a	(+/-)-12b	$(+)-1^{b}$	$(+)-2^{b}$	(+/-)-5 ^c	(+/-)-2-epi-2 ^a	(+/-)-12c
%GI (at 100 μ M) IC ₅₀ (μ g mL ⁻¹)	85% 1.57	3.8% >30	7% >30	 0.048	 0.29	 2.8	16% >30	3.5% >30
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^{*a*} From ref. 28. ^{*b*} From ref. 17. ^{*c*} From ref. 19*b*

discussion of the role of aromatic substituents in the pharmacophore of pancratistatin.

Experimental section

(1*S**,5*R**,6*R**,7*R**,8*S**)-6-(Furan-2-yl)-8-((2-methoxypropan-2-yl)oxy)-3,3-dimethyl-7-nitro-2,4-dioxabicyclo[3.3.1]nonan-9-one (+/-)-22

2-Methoxypropene (966 µL, 10.09 mmol) and PPTS (84 mg, 0.34 mmol) were added to a solution of (+/-)-15 (1 g, 3.36 mmol) in dry CH₂Cl₂ (16.8 mL). After stirring for 1 h at rt, the reaction mixture was neutralized with Et₃N (0.5 mL) and the solvent evaporated in vacuo. Chromatography (15% EtOAchexane) afforded 22 (1.22 g, 97%) as a white solid: $R_{\rm f} = 0.49$ (20% EtOAc-hexane); mp: 137–139 °C (EtOAc-hexane); ¹H NMR (CDCl₃, 400 MHz) δ : 7.37 (dd, *J* = 1.8 Hz, 0.7 Hz, 1H), 6.41 (d, J = 3.3 Hz, 1H), 6.35 (dd, J = 3.3, 1.8 Hz, 1H), 5.45 (dd, J = 11.7, 9.6 Hz, 1H), 4.69 (dd, J = 1.9, ≈1.9 Hz, 1H), 4.45 (dd, *J* = 2.2, ≈2.2 Hz, 1H), 4.30 (dd, *J* = 9.6, 1.9 Hz, 1H), 3.51 (dd, *J* = 11.7, 1.9 Hz, 1H), 3.24 (s, 3H), 1.57 (s, 3H), 1.47 (s, 3H), 1.37 (s, 3H), 1.30 (s, 3H); $^{13}\mathrm{C}$ NMR (CDCl_3, 100 MHz) δ : 206.7, 147.6, 143.0, 110.7, 108.9, 102.8, 99.3, 86.9, 79.1, 76.7, 76.5, 49.7, 44.4, 28.3, 25.3, 24.9, 24.1; LRMS (IE) m/z (%): 369.0 [(M)⁺, 8]; HRMS [IE, (M)⁺] m/z: calcd for (C₁₇H₂₃NO₈): 369.1424, found: 369.1414.

(1*S**,5*R**,6*R**,7*R**,8*S**)-7-Amino-6-(furan-2-yl)-8-((2-methoxypropan-2-yl)oxy)-3,3-dimethyl-2,4-dioxabicyclo[3.3.1]-nonan-9-ona (+/-)-23

A suspension of 22 (3.52 g, 9.52 mmol) and RANEY®-Nickel (≈15 mL) in MeOH-THF (1:1, 40 mL) was stirred at rt under a H_2 atmosphere. After completion of the reduction (as monitored by TLC), the catalyst was filtered off and washed with MeOH (200 mL) and EtOAc (200 mL). Evaporation of the combined washings and the filtrate afforded 23 (2.76 g, 86%) as a pale brown solid; $R_{\rm f} = 0.42$ (80% EtOAc-hexane): mp = 137-139 °C (EtOAc-hexane); ¹H NMR (CDCl₃, 400 MHz) δ: 7.35 (d, J = 1.7 Hz, 1H), 6.35 (dd, J = 3.2, 1.7 Hz, 1H), 6.32 (d, J = 3.2)3.2 Hz, 1H), 4.60 (dd, J = 2.3, 1.8 Hz, 1H), 4.25 (dd, J = 2.3, 1.9 Hz, 1H), 3.89 (dd, J = 11.0, 9.2 Hz, 1H), 3.51 (dd, J = 9.2, 1.9 Hz, 1H), 3.27 (s, 3H), 2.66 (dd, J = 11.0, 1.8 Hz, 1H), 1.51 (s, 3H), 1.42 (s, 9H); 13 C NMR (CDCl₃, 100 MHz) δ : 209.4, 152.0, 142.0, 110.6, 107.6, 102.2, 98.8, 80.2, 79.7, 77.9, 50.0, 49.6, 48.0, 28.4, 25.4, 25.2, 25.2; LRMS (EI) m/z (%): 340.1 $[(M + H)^+, 12].$

(1*R**,5*S**,6*S**,7*R**,8*R**)-7-Acrylamido-8-(furan-2-yl)-3,3dimethyl-9-oxo-2,4-dioxabicyclo[3.3.1]nonan-6-yl methyl carbonate (+/-)-16c

Acryloyl chloride (400 μ L, 4.99 mmol), Et₃N (696 μ L, 4.99 mmol) and DMAP (110 mg, 0.91 mmol) were added to a solution of 23 (1.54 g, 4.54 mmol) in dry CH₂Cl₂ (22 mL) under argon. After stirring for 2 h, 2,6-di-*tert*-butyl-4-methyl-phenol (200 mg, 0.91 mmol) was added, the solvent was evaporated and the residue dissolved in dry MeOH (30 mL). PPTS

(228 mg, 0.91 mmol) was added and the mixture stirred at rt for 1 h and then neutralized with Et₃N. After solvent evaporation, the residue was dissolved in dry CH₂Cl₂ (30 mL) and treated with methyl chloroformate (420 µL, 5.44 mmol) and DMAP (664 mg, 5.44 mmol). After stirring for 1.5 h, 2,6-di-tertbutyl-4-methylphenol (200 mg, 0.91 mmol) was added and the mixture treated with a saturated aqueous solution of NaHCO3 (30 mL) and extracted with CH_2Cl_2 (3 × 20 mL). Chromatography (40% EtOAc-hexane) afforded 16c (1.24 g, 72%) as a white solid: $R_{\rm f} = 0.67$ (60% EtOAc-hexane); ¹H NMR (CDCl₃, 300 MHz) δ: 7.33 (s, 1H), 6.44 (d, J = 3.2 Hz, 1H), 6.37–6.29 (m, 1H), 6.17 (dd, J = 17.0, 1.3 Hz, 1H), 5.95 (dd, J = 17.0, 10.2 Hz, 1H), 5.82 (d, J = 8.8 Hz, 1H), 5.58 (dd, J = 10.2, 1.3 Hz, 1H), 5.12 (ddd, J = 11.8, 10.3, 8.8 Hz, 1H), 4.94 (dd, J = 10.3, 2.1 Hz, 1H), 4.68 (s, 1H), 4.44 (s, 1H), 3.76 (s, 3H), 3.32 (br d, J = 11.8 Hz, 1H), 1.59 (s, 3H), 1.48 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ: 206.4, 165.7, 154.9, 150.0, 142.1, 130.4, 127.0, 110.8, 108.5, 99.4, 79.1, 77.7, 77.5, 55.3, 48.6, 44.5, 28.2, 25.4; LRMS (CI) m/z (%): 380.1 [(M + H)⁺, 100]; HRMS [CI, (M + H)⁺] *m*/*z*: calcd for (C₁₈H₂₂NO₈): 380.1345, found: 380.1340.

(1*R**,5*S**,6*S**,7*R**,8*R**)-7-Acrylamido-8-(5-chlorofuran-2-yl)-3,3dimethyl-9-oxo-2,4-dioxabicyclo[3.3.1]nonan-6-yl methyl carbonate (+/-)-16d

N-Chlorosuccinimide (262 mg, 1.96 mmol) was added to a solution of 16c (620 mg, 1.63 mmol) in dry DMF (8 mL) under argon. After stirring for 12 h at rt, the reaction mixture was neutralized with Et₃N, treated with 2,6-di-tert-butyl-4-methylphenol (72 mg, 0.33 mmol) and the solvent was evaporated in vacuo. Chromatography (40% EtOAc-hexane) afforded 16d (640 mg, 95%) as a white solid: $R_{\rm f} = 0.51$ (50% EtOAc-hexane); mp: 139–145 °C (CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ: 6.46 (d, J = 3.3 Hz, 1H), 6.20 (dd, J = 17.0, 1.3 Hz, 1H), 6.09 (d, J =3.3 Hz, 1H), 5.97 (dd, J = 17.0, 10.3 Hz, 1H), 5.79–5.65 (m, 1H), 5.62 (dd, J = 10.3, 1.3 Hz, 1H), 5.10 (ddd, J = 11.7, 10.3, 9.8 Hz, 1H), 4.90 (dd, J = 10.2, 2.0 Hz, 1H), 4.67 (dd, J = 2.0, 2.0 Hz, 1H), 4.43 (dd, J = 2.0, 1.3 Hz, 1H), 3.76 (s, 3H), 3.24 (dd, J = 11.7, 1.3 Hz, 1H), 1.59 (s, 3H), 1.48 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ: 206.2, 166.0, 154.8, 149.6, 135.3, 130.4, 127.0, 110.9, 107.4, 99.3, 79.4, 77.3, 77.2, 55.2, 47.5, 44.6, 28.1, 25.3; LRMS (CI) m/z (%): 414.1 [(M + H)⁺, 22], 356.0 (37), 280.0 (46), 267.0 (95); HRMS [CI, $(M + H)^+$] m/z: calcd for $(C_{18}H_{21}ClNO_8)$: 414.0956, found: 414.0956.

Oxanorbornene (+/-)-18a

A suspension of **16d** (80 mg, 0.19 mmol) and NaHCO₃ (19 mg, 0.23 mmol) in xylenes (2 mL) was stirred in a closed glass tube for 5 h at 140 °C (external bath temperature). Rotary evaporation of the solvent and chromatography (40% EtOAc–hexane) afforded **18a** (42 mg, 53%) as a white solid: $R_{\rm f}$ = 0.57 (50% EtOAc–hexane); mp = 143–145 °C (EtOAc–hexane, decomposition); ¹H NMR (CDCl₃, 300 MHz) δ : 7.62 (d, J = 6.9 Hz, 1H), 6.78 (d, J = 5.6 Hz, 1H), 6.38 (d, J = 5.6 Hz, 1H), 4.65 (dd, J = 2.3, 2.3 Hz, 1H), 4.54 (dd, J = 9.9, 2.3 Hz, 1H), 4.49 (br s, 1H), 4.34 (ddd, J = 11.5, 9.9, 6.9 Hz, 1H), 3.80 (s, 3H), 2.65 (dd, J = 11.8, 4.0 Hz, 1H), 2.49 (dd, J = 8.3, 4.0 Hz, 1H), 2.11 (dd, J =

11.8, 8.3 Hz, 1H), 1.91 (dd, J = 11.5, 2.3 Hz, 1H), 1.48 (s, 3H), 1.44 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ : 204.7, 171.7, 154.9, 140.9, 136.8, 99.7, 98.5, 88.4, 79.6, 77.0, 75.9, 55.9, 49.0, 48.2, 47.5, 38.2, 28.7, 25.6; LRMS (CI) m/z (%): 414.14 [(M + H)⁺, 10], 267.0 (15), 56.0 (100); HRMS [CI, (M + H)⁺] m/z: calcd for (C₁₈H₂₁ClNO₈): 414.0956, found: 414.0950.

Triacetate (+/-)-18b

A mixture of 18a (118.4 mg, 0.29 mmol) and Dowex 50WX (438 mg) in MeOH (2.9 mL) was stirred for 24 h at 60 °C. After filtration, the solvent was evaporated in vacuo and the crude dissolved in DCE-THF (1:1, 2.9 mL) under argon. Na(AcO)₃BH (303.7 mg, 1.45 mmol) was added and the mixture stirred at rt for 2 h and then quenched with 30% aqueous hydrogen peroxide (0.8 mL). After solvent evaporation, the crude was dissolved in dry CH₂Cl₂ (2.9 mL) and treated with Et₃N (1.2 mL), Ac₂O (0.6 mL) and DMAP (6.8 mg, 0.06 mmol). After stirring for 12 h at rt, the mixture was treated with a saturated aqueous solution of NaHCO₃ (2.9 mL) and extracted with CH_2Cl_2 (3 × 2 mL). Chromatography (60% EtOAc-hexane) gave 18b (121.8 mg, 85%) as a white foam: $R_{\rm f} = (20\% \text{ EtOAc-hexane}); {}^{1}\text{H}$ NMR (CDCl₃, 400 MHz) δ : 7.41 (s, 1H), 6.50 (d, J = 5.7 Hz, 1H), 6.32 (d, J = 5.7 Hz, 1H), 5.53 (dd, J = 3.7, 2.9 Hz, 1H), 5.30 (dd, J = 2.9, 2.9 Hz, 1H), 5.14 (br s, 1H), 4.89 (dd, J = 10.6, 3.7 Hz, 1H), 4.20 (dd, J = 11.9, 10.6 Hz, 1H), 3.84 (s, 3H), 2.80 (dd, J = 12.0, 3.8 Hz, 1H), 2.65 (dd, J = 11.9, 2.3 Hz, 1H), 2.54 (dd, J = 8.3, 3.8 Hz, 1H), 2.22 (dd, J = 12.0, 8.3 Hz, 1H), 2.18 (s, 3H), 2.14 (s, 3H), 2.11 (s, 3H); $^{13}\mathrm{C}$ NMR (CDCl_3, 100 MHz) δ : 171.1, 168.9, 168.8, 168.1, 154.4, 141.8, 134.6, 97.7, 88.2, 75.3, 69.2, 66.9, 66.5, 55.7, 47.7, 46.7, 41.4, 38.8, 21.0, 20.7, 20.6; LRMS (CI) m/z (%): 502.1 [(M + H)⁺, 22], 460.1 (16), 123.1 (18); HRMS $[ICI, (M + H)^{+}] m/z$: calcd for $(C_{21}H_{25}ClNO_{11})$: 502.1116, found: 502.1110.

7,9-Dideoxy-8-hydroxypancratistatin (+/-)-12a

A solution of **18b** (84 mg, 0.17 mmol) in MeOH (1.6 mL) was treated with NaMeO in MeOH (5.4 M, 186 μ L) and stirred for 6 h at 50 °C. The pH of the mixture was adjusted to 3 with a 1 M aqueous solution of HCl and the solvent evaporated *in vacuo*. Chromatography (20% MeOH–CH₂Cl₂–1% TFA) afforded **12a** (14 mg, 93%) as a white solid: $R_{\rm f}$ = 0.28 (20% MeOH–CH₂Cl₂–1% TFA); ¹H NMR (CDCl₃, 400 MHz) δ : 7.42 (d, J = 2.7 Hz, 1H), 7.27 (d, J = 8.4 Hz, 1H), 7.00 (dd, J = 8.4, 2.7 Hz, 1H), 4.52 (br s, 1H), 4.18 (dd, J = 3.2, 3.2 Hz, 1H), 4.02 (br s, 1H), 3.93–3.88 (m, 2H), 3.21–314 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ : 168.4, 157.4, 140.9, 131.3, 127.4, 121.1, 115.3, 75.2, 72.2, 72.0, 70.4, 51.9, 40.8; LRMS (CI) *m/z* (%): 282.0 [(M + H)⁺, 100], 203.0 (100), 167.0 (37); HRMS [CI, (M + H)⁺] *m/z*: calcd for (C₁₃H₁₆NO₆): 282.0978, found: 282.0972.

7,9-Dideoxy-8-[2-(benzyloxy)-2-oxoethoxy]pancratistatin (+/-)-12b

 K_2CO_3 (30 mg, 0.17 mmol), benzyl bromoacetate (21 μ L, 0.13 mmol) and $Bu_4N^+I^-$ (12 mg, 0.03 mmol) were added to a solution of **12a** (23 mg, 0.08 mmol) in dry DMF (1 mL).

After stirring for 9 h at rt, the pH was adjusted to 3 with 1 M aqueous HCl and the solvent evaporated *in vacuo*. Chromatography (20% MeOH–CH₂Cl₂–1% TFA) gave **12b** (9.1 mg, 25%) as a white solid: $R_{\rm f}$ = 0.62 (20% EtOAc–hexane); ¹H NMR (CDCl₃, 300 MHz) δ : 7.51 (d, J = 2.8 Hz, 1H), 7.38 (d, J = 8.6 Hz, 1H), 7.35–7.22 (m, 5H), 7.16 (dd, J = 8.6, 2.8 Hz, 1H), 4.76 (s, 2H), 4.59 (s, 2H), 4.53 (br s, 1H), 4.18 (dd, J = 3.2, 3.2 Hz, 1H), 4.02 (br s, 1H), 3.96–3.87 (m, 2H), 3.26–3.17 (m, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ : 170.0, 166.5, 157.0, 141.5, 132.3, 130.5, 128.1 (2C), 127.1, 126.8 (2C), 126.3, 119.7, 112.5, 74.0, 70.9, 70.8, 69.1, 64.9, 64.0, 50.6, 39.7.

7,9-Dideoxy-2-epi-8-hydroxypancratistatin (+/-)-12c

A mixture of 18a (45 mg, 0.11 mmol) and NaBH₄ (8 mg, 0.22 mmol) in dry MeOH (0.7 mL) was stirred for 10 min at rt. The reaction was neutralized with 5% aqueous AcOH, evaporated in vacuo and extracted with CH_2Cl_2 (3 × 0.5 mL). The residue was dissolved in CH₂Cl₂ (0.5 mL) and stirred with p-TsOH·H₂O (25 mg, 0.13 mmol) for 1 h at rt. After neutralization with Et₃N and evaporation of the volatiles in vacuo, dry CH₂Cl₂ (0.5 mL), Et₃N (121 µL, 0.87 mmol), Ac₂O (41 µL, 0.43 mmol) and DMAP (3 mg, 0.022 mmol) were added and the mixture stirred for 1 h at rt, diluted with a saturated aqueous solution of NH₄Cl (0.5 mL) and extracted with CH_2Cl_2 (3 × 0.5 mL). Chromatography (60% EtOAchexane) gave protected 12c [27 mg, 50% for the three steps, $R_{\rm f} = 0.24$ (70% EtOAc-hexane)], which was dissolved in MeOH (1 mL), treated with NaMeO (5.4 M in MeOH, 60 µL) and stirred for 6 h at 50 °C. pH adjustment to 3 with aqueous HCl (1 M), solvent evaporation in vacuo and chromatography (20% MeOH-CH₂Cl₂-1% TFA) afforded 12c (13 mg, 93% for the last step) as a white solid: $R_{\rm f} = 0.25$ (20% MeOH-CH₂Cl₂-1% TFA); ¹H NMR (CDCl₃, 400 MHz) δ : 7.39 (d, J = 2.6 Hz, 1H), 7.26 (d, J = 8.4 Hz, 1H), 6.98 (dd, J = 8.4, 2.6 Hz, 1H), 4.61 (s, 1H), 4.09 (s, 1H), 3.83 (dd, J = 13.3, 10.2 Hz, 1H), 3.68 (br s, 1H), 3.59 (dd, J = 10.2, 2.6 Hz, 1H), 2.78 (br d, J = 13.3 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ : 168.1, 157.6, 131.4, 130.3, 127.5, 121.0, 115.2, 76.3, 73.9, 70.8, 51.5, 42.5; LRMS (ESI-TOF) m/z (%): 304.1 [(M + Na)⁺ 36], 282.1 $[(M + H)^+$, 58], 245.1 (100), 149.0 (49); HRMS [ESI-TOF, $(M + H)^+$] m/z: calcd for $(C_{13}H_{16}NO_6)$: 282.0978, found: 282.0972.

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